PRELIMINARY SCHEDULE CRISPR/Cas9 workshop at SNU, May 24-28 2018

Theory = 9:00 to 12:30 Practical = 14:00 to 18:00

Day 1: May 24

Theory: Introduction to Genome Editing; CRISPR/Cas9-based Editing – NHEJ and HDR; Complete Workflow for Genome Editing

Practical:

> Basic outline of the practical training planned for the 5 days (cloning, transfection & overview of CRISPR strategy);

> Start gDNA cloning – annealing of gDNA oligos, ligation into pre-cut pDC2-Cas9 vector, transformation of ligation mix into bacteria.

>> P. falciparum ring-stage transfection (preexisting construct)

Day 2: May 25

Theory: Applications of CRISPR/Cas9 Technologies – Beyond Genome Editing

Practical:

> Start screening of gDNA clones (inoculate for plasmid prep)

>> Transfection of preexisting gDNA/Cas9 construct (OFP-expressing and cloned using ThermoFisher's GeneArt CRISPR Nuclease Vector with OFP Reporter Kit) into HEK293 cells using nucleofection

>> <u>If time permits:</u> Transfection of gRNA-Cas9 ribonucleoprotein complex (pre-assembled using ThermoFisher's GeneArt[™] Precision gRNA and Cas9 Platinum nuclease) with lipofectamine

Day 3: May 26

Theory: Live projects of gDNA design, discussion of downstream steps, project-specific discussions (pre-defined for workshop participants), etc.

Practical:

- > Plasmid prep and PCR-based detection of gDNA positive clones
- > Cloning of Homology boxes 1 and 2 using GIBSON assembly,

transformation of GIBSON assembly mix into bacteria

>> GIEMSA staining of transfected P. falciparum parasites

>> Observe transfected cells for OFP signal

Day 4: May 27

Theory: CRISPR/Cas9 advances in pathogenic parasites; selected 15 min talks from 4 workshop participants

Practical:

Start screening of homology box clones (inoculate for plasmid prep)
Observe transfected cells for OFP signal; Harvest the transfected
HEK293 cells, genomic DNA preparation, confirmation of deletion (using
ThermoFisher's GeneArt GenomicCleavage Detect Kit)

>> <u>If time permits:</u> Harvest cells transfected with gRNA-Cas9 ribonucleoprotein complex and analyze as described above.

Day 5: May 28

Theory: ThermoFisher CRISPR/Cas9 genome-editing resources

Practical:

> Confirm homology box clones using restriction digestion of plasmids, Results analysis...

>> GIEMSA staining of transfected P. falciparum parasites

>> Results analysis and discussion...